Thus, the biosynthesis of the carbocyclic acid. rings of phenazine pigments are, in all probability, additional cases of aromatic biosynthesis proceeding via the shikimic acid pathway.10

Anthranilic acid-1-14C was prepared by the method of Murray and Ronzio<sup>11</sup> and the labeled substrate was fed over a period of five days to a culture of Pseudomonas chlororaphis NRRL B-977<sup>11a</sup> growing on a peptone-glycerol broth<sup>4</sup> medium.

The pigment was isolated by ether extraction and purified by chromatography on alumina and crystallization from methanol. During this purification the original chlororaphin is entirely oxidized to phenazine-1-carboxamide, which is obtained as bright yellow needles, m.p. 241.5-243 (lit.<sup>12</sup> 241°).

The phenazine-1-carboxamide was hydrolyzed in base to the corresponding acid. The acid so obtained had, in all cases within the limits of experimental error, the same activity as the amide that had itself been purified to constant activity, thus rendering improbable the possibility that the observed activity was due to some highly active minor impurity.

Heating phenazine-1-carboxylic acid in diphenvl ether with copper powder at 260° for five hours resulted in decarboxylation and yielded phenazine, which was purified by chromatography over alumina followed by recrystallization from ethanol to give needles, m.p. 171-171.5° (lit. 171°).12

Similar degradations were carried out on chlororaphin obtained from a culture medium to which had been added sodium carbonate-14C, alanine-1-14C and alanine-2-14C. The specific activities of the various samples were determined either by combustion to carbon dioxide, which was counted in an ion chamber by means of a vibrating reed electrometer, or by counting the samples directly in a Nuclear Chicago gas-flow counter, model D47. The results of these experiments are shown in Table I.

Substrate	TABLE I % Incorporation	% Activity in phenazine ring <sup>a</sup>
Anthranilic acid car-	0.002	$21 \pm 3\%$
Sodium carbonate-14C	0.0001	$60 \pm 10\%$
Alanine-1-14C	0.01	$25 \pm 4\%$
Alanine-2-14C	0.01	$71 \pm 2\%$

<sup>a</sup> The remaining activity was in all cases found in the carbon dioxide, collected as barium carbonate from the decarboxylation.

Thus, the incorporation of anthranilic acid carboxyl-14C into chlororaphin has been demonstrated. The fact that all of the activity so incorporated does not reside in the carboxyl carbon of the phenazine pigment requires comment. It is known that anthranilic acid is very rapidly metabolized<sup>13</sup> by Pseudomonas species with loss of the carboxyl carbon as carbon dioxide and formation of catechol.14 It is

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(14) J. C. Yanofsky in "Amino Acid Metabolism," Editors W. D. McElroy and H. B. Glass, Johns Hopkins Press, Baltimore, Md., 1959, pp. 930-939

likely that some of the carbon dioxide resulting from this metabolic decarboxylation is incorporated by diverse pathways into precursors of the phenazine pigments. We propose that <sup>14</sup>C proceeding by such pathways accounts for the activity found in the aromatic ring system of chlororaphin biosynthesized from anthranilic acid carboxyl-14C as substrate; *i.e.*, the experimentally observed distribution of activity represents the sum of intact anthranilic acid incorporation and a more circuitous incorporation of the carboxyl carbon of anthranilic acid via a pathway that includes carbonate. The results with added sodium carbonate-14C support this suggestion.

The distributions of activity found in chlororaphin produced with alanine-1-14C and alanine-2-14C as labeled substrates are also in accord with their incorporation via a pathway involving shikimic acid and anthranilic acid.15

It is possible to write several detailed mechanisms for the dimerization reaction, some of which involve preliminary oxidation at nitrogen, or prior conversion of one of the rings to 3-hydroxyanthranilic acid. Since the precise structures of the species that couple is not defined by the experiments reported herein, the selection of one particular mechanism is premature. Further, it should be emphasized that possibilities are not limited to a dimerization of two identical units. These experiments do indicate, however, that the ring in the phenazine pigment bearing the carboxyl carbon has not proceeded via 3-hydroxyanthranilic acid as this pathway, which includes tryptophan, would result in loss of the carboxyl carbon originally present in the anthranilic acid substrate.<sup>16</sup>

(15) A. White, P. Handler, E. L. Smith and DeWitt Stetten, Jr., "Principles of Biochemistry," 2nd Ed., McGraw-Hill Book Co., New

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**Contribution No. 2655** R. E. CARTER GATES AND CRELLIN LABORATORIES OF CHEMISTRY

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## ON THE IONIZATION OF POLYSTYRENE SULFONIC ACID

## Sir:

Raman spectroscopy has proven to be one of the most useful techniques for the determination of the extent of ionization of ordinary electrolytes.1 In this note we communicate the results of preliminary Raman spectroscopic studies of the state of ionization of a strong polyelectrolyte with the intention of clarifying the nature of ion-binding, a phenomenon commonly encountered with these substances.

We have obtained the Raman spectra of polystyrenesulfonic acid (PSSA) and its monomeric unit, p-ethylbenzenesulfonic acid (EBSA) using a three prism Steinheil spectrograph with photoelectric recording.<sup>2</sup> The spectra were excited by the

(1) T. F. Young, L. F. Maranville and H. M. Smith, "The Structure of Electrolytic Solutions," W. J. Hamer, Editor, J. Wiley and Sons. Inc., New York, N. Y., 1959, Chapter 4 and the references contained therein.

(2) T. F. Young, Record Chem. Prog., 12, 81 (1951).

4358 Å. line from high intensity mercury lamps. PSSA was obtained from Dr. W. N. Vanderkoi of the Dow Chemical Co., Midland, Michigan. The molecular weight was approximately 10<sup>6</sup>. The original sample was purified by solution in water, precipitation with isopropyl alcohol in a Waring Blendor and thereafter passage through a column containing cation exchange resin (Dowex 50). EBSA was synthesized from ethylbenzene and sulfuric acid and purified by standard methods.

PSSA was studied in aqueous solutions at a concentration of 0.15 N and EBSA was studied at a concentration of 1.0 N. It would have been advantageous to study PSSA in more concentrated solution, but this proved impossible because of excessive light scattering. Because of the low concentration of the solutions of PSSA the number of Raman lines detected in these solutions is fewer than that observed in solutions of EBSA and the intensities are also lower. Because of intense light scattering, no lines closer to the exciting line than 965 cm.<sup>-1</sup> could be observed in solutions of PSSA. The results are recorded in Table I.

TABLE I

RAMAN	SPECTRA OF PSSA AND	EBSA (cm. <sup>-1</sup> )
EBSA	PSSA	
620 (w)		
780 (s)		
965 (w)	965 (vw)	
1035 (w)	1030 (vw)	
1060 (w)	1060 ( <b>vw</b> )	
1125 (vs)	1125 (s)	w, weak
1205 (w)	• • • •	s, strong
1450 (w)	1450 (vw)	vw, very weak
1590 (vs)	1590 (s)	vs, very strong

It is clear that the lines appearing in both spectra are identical. The 1125 cm.<sup>-1</sup> line has been identified by Nisi<sup>3</sup> as characteristic of the ionized sulfonic acid residue, whereas the other strong lines arise from the vibrational modes of the *p*-substituted benzene nucleus and are commonly observed in these compounds.<sup>4</sup>

Mock and Marshall have shown that solutions of PSSA have very low apparent degrees of ionization.<sup>5,6</sup> They have found a degree of ionization of 0.38 which is independent of concentration. The absence of any new lines in the spectrum of PSSA and the observation that the relative intensities of the lines observed are the same as in EBSA strongly suggests that PSSA is completely ionized. This conclusion is in complete accord with nuclear magnetic resonance measurements of the proton shift in solutions of PSSA made in this laboratory.<sup>7</sup> It is clear, then, that the low apparent degree of ionization of PSSA is due to electrostatic interaction between the hydrogen ions and the polyion. Although the counterions are localized in a small region near the charged polymer skeleton, they do not associate at the skeletal sites as if they were weak electrolytes. The counterions must be as-

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(6) R. A. Mock, C. A. Marshall and T. E. Slykhouse, J. Phys. Chem., 58, 498 (1954).

sumed to form a "mobile monolayer" about the polyion skeleton with the very strong electrostatic interactions possible localizing individual counterions in what may be thought of as a quasi-cell in the vicinity of a fixed skeletal charge, but not "on the charge."<sup>8</sup>

We wish to thank the United States Public Health Service, the Air Force Research and Development Command and the National Science Foundation for financial support. We also wish to thank Prof. T. F. Young for the use of the Raman spectrograph and Mr. Don Irish for assistance with the measurements.

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(9) Department of Chemistry, University of Ljubljana, Ljubljana, Yugoslavia.

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RECEIVED DECEMBER 7,	1960

## IRON TRICARBONYL COMPLEXES OF CYCLOHEPTATRIENE, CYCLOHEPTADIENE AND CYCLOHEPTADIENIUM ION<sup>1</sup>

## Sir:

Hydride ion abstraction from  $\pi$ -cycloheptatrienemetal-tricarbonyls (I) by triphenylcarbonium ion has been shown to provide a common route to the  $\pi$ -tropenium ion complexes (II),  $(\pi$ -C<sub>7</sub>H<sub>7</sub><sup>+</sup>)M-(CO)<sub>3</sub>,BF<sub>4</sub><sup>-</sup> where M = Mo,<sup>2a</sup> Cr<sup>2b,3</sup> and W.<sup>2b</sup> Recently, preliminary undetailed reports of extension of this method by Fischer and Palm<sup>4</sup> and by Pauson<sup>5</sup> to the  $\pi$ -cycloheptatriene-iron carbonyl complex, presumed to be  $(\pi-C_7H_8)Fe(CO)_2$  by Wilkinson and co-workers<sup>6</sup> and by Manuel and Stone,<sup>7</sup> have contended that the product is a  $\pi$ tropenium ion complex, either  $(\pi - C_7 H_7^+) Fe(CO)_2^4$ or  $(\pi - C_7 H_7^+) Fe(CO)_3.^5$  We wish to report our substantially different results which clearly demonstrate that cycloheptatriene reacts with iron pentacarbonyl to give iron tricarbonyl complexes of cycloheptatriene (I,  $M = Fe)^5$  and, more slowly, of cycloheptadiene (III), that triphenylcarbonium fluoroborate shows addition rather than hydride ion abstraction with the cycloheptatriene complex (I, M = Fe) to yield the trityl adduct (IV, R =  $(C_6H_5)_3C-$ ), and that the stable cycloheptadienium ion complex (IV, R = H)<sup>8</sup> is formed readily either by proton addition to the cycloheptatriene complex

(1) Partial support of this work by the Office of Ordnance Research, U. S. Army, is gratefully acknowledged.

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<sup>(4)</sup> N. S. Ham and A. H. Hambly, Aust. J. Chem., 6, 135 (1953).

<sup>(7)</sup> L. Kotin and M. Nagasawa, J. Am. Chem. Soc., in press.